

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/340498426>

Clinical and Bacterial Markers of Periodontitis and Their Association with Incident All-Cause and Alzheimer's Disease Dementia in a Large National Survey

Article in *Journal of Alzheimer's disease: JAD* · April 2020

DOI: 10.3233/JAD-200064

CITATIONS

92

READS

1,679

6 authors, including:



May A Beydoun

National Institute on Aging

242 PUBLICATIONS 13,788 CITATIONS

[SEE PROFILE](#)



Hind Beydoun

U.S. Department of Veterans Affairs

288 PUBLICATIONS 7,546 CITATIONS

[SEE PROFILE](#)



Alan Zonderman

National Institute on Aging

707 PUBLICATIONS 37,691 CITATIONS

[SEE PROFILE](#)

Clinical and Bacterial Markers of Periodontitis and Their Association with Incident All-Cause and Alzheimer's Disease Dementia in a Large National Survey

May A. Beydoun^{a,*1}, Hind A. Beydoun^b, Sharmin Hossain^a, Ziad W. El-Hajj^c, Jordan Weiss^d and Alan B. Zonderman^a

^aLaboratory of Epidemiology and Population Sciences, National Institutes on Aging, NIA/NIH/IRP, Baltimore, MD, USA

^bFort Belvoir Community Hospital, Fort Belvoir, VA, USA

^cMcGill University, Montreal, QC, Canada

^dPopulation Studies Center and the Leonard Davis Institute of Health Economics, University of Pennsylvania, Philadelphia, PA, USA

Accepted 19 February 2020

Abstract. Microbial agents including periodontal pathogens have recently appeared as important actors in Alzheimer's disease (AD) pathology. We examined associations of clinical periodontal and bacterial parameters with incident all-cause and AD dementia as well as AD mortality among US middle-aged and older adults. Clinical [Attachment Loss (AL); probing pocket depth (PPD)] and bacterial [pathogen immunoglobulin G (IgG)] periodontal markers were investigated in relation to AD and all-cause dementia incidence and to AD mortality, using data from the third National Health and Nutrition Examination Surveys (NHANES III, 1988–1994) linked longitudinally with National Death Index and Medicare data through January 1, 2014, with up to 26 years of follow-up. Sex- and age-specific multivariable-adjusted Cox proportional hazards models were conducted. Among those ≥ 65 years, AD incidence and mortality were consistently associated with PPD, two factors and one cluster comprised of IgG titers against *Porphyromonas gingivalis* (*P. gingivalis*), *Prevotella melaninogenica* (*P. melaninogenica*) and *Campylobacter rectus* (*C. rectus*) among others. Specifically, AD incidence was linked to a composite of *C. rectus* and *P. gingivalis* titers (per SD, aHR=1.22; 95% CI, 1.04–1.43, $p=0.012$), while AD mortality risk was increased with another composite (per SD, aHR=1.46; 95% CI, 1.09–1.96, $p=0.017$) loading highly on IgG for *P. gingivalis*, *Prevotella intermedia*, *Prevotella nigrescens*, *Fusobacterium nucleatum*, *C. rectus*, *Streptococcus intermedius*, *Capnocytophaga Ochracea*, and *P. melaninogenica*. This study provides evidence for an association between periodontal pathogens and AD, which was stronger for older adults. Effectiveness of periodontal pathogen treatment on reducing sequelae of neurodegeneration should be tested in randomized controlled trials.

Keywords: Aging, Alzheimer's disease, dementia, periodontal pathogens, periodontitis

INTRODUCTION

Dementia, a common disorder affecting older adults, has an estimated prevalence of 4.7% (≥ 60 years) [1], with 4.6–7.7 million additional annual cases occurring worldwide [3.5–10.5 per 1,000 in various world regions] [1–3]. Generally, ~60–80%

*Correspondence to: May A. Beydoun, PhD, NIH Biomedical Research Center, National Institute on Aging, IRP, 251 Bayview Blvd., Suite 100, Room #: 04B118, Baltimore, MD 21224, USA. Fax: +1 410 558 8236; E-mail: baydounm@mail.nih.gov.

¹MAB had full access to the data used in this manuscript and completed all the statistical analyses.

of dementia is ascribed to Alzheimer's disease (AD) [1], a progressive neurodegenerative disorder with a multi-factorial etiology. AD triggers progressive episodic memory deterioration followed by impairment in other domains of cognition [4]. AD is likely caused by age-dependent and progressive A β -amyloid brain deposition [5], with a second pathological hallmark being the neurofibrillary tangles arising from hyperphosphorylated tau proteins [6]. It constitutes the primary cause of disability among older adults [7], the leading health care burden in developed countries [8], and the sixth leading cause of death in the US [9]. The number of AD-affected Americans is expected to rise from currently 5.4 million to 13.8 million by 2050 [9]. In 2016, US long-term and hospice care cost for all-cause dementia (including AD, vascular dementia and other rare forms) was estimated at \$236 billion [9].

Despite no effective treatment, epidemiologic research has uncovered genetic markers for late-onset AD (e.g., ApoE ϵ 4) and several modifiable risk factors. The combined effects of low education, smoking, physical inactivity, depression, mid-life obesity, hypertension, and type 2 diabetes explains ~54% of AD risk [10], leaving much variation unaccounted for. Identifying novel mid-life modifiable risk factors is essential for planning cost-effective interventions. Microbial agents have recently appeared as important actors of AD's etiology [8], notably periodontal pathogens [11–15], many of which can cause periodontitis (Pd), a condition shown to increase risk of diabetes, atherosclerosis, cardiovascular events [16], and adverse cognitive outcomes [11–15].

Pd affects 20–50% of older adults and is initiated by periodontal bacteria, the most well-known being *Porphyromonas gingivalis* (*P. gingivais*), *Tannerella forsythia* (*T. forsythia*), *Aggregatibacter actinomycetemcomitans* (*A. actinomycetemcomitans*), and *Treponema denticola* (*T. denticola*), triggering gingival inflammation, connective tissue destruction, periodontal pocket formation, alveolar bone loss, and edentulation [17]. Given its increased prevalence with age, Pd may be highly predictive of AD. In fact, several hypothesized pathways link Pd to AD, including brain tissue invasion by periodontal gram-negative bacteria found in the dental biofilm, release of bacterial byproducts into the brain via bloodstream invasion and direct impacts of peripheral nerves [18]. Periodontal pathogens can affect brain cytokines through systemic or neural pathways [19]. A recent comprehensive study suggests that *P. gingi-*

valis and its associated gingipains in the brain play a central role in AD pathogenesis and suggests that A β ₁₋₄₂ is produced in the brain partly as a response to this infection [20]. However, the epidemiological evidence as to the relationship between various Pd-related pathogens, including *P. gingivalis*, and AD remains scarce.

We examined age and sex-specific associations of serum immunoglobulin G (IgG) humoral immune response against periodontal pathogens and Pd markers with incident all-cause and AD dementia as well as AD mortality among U.S. middle-aged and older adults (45+years at baseline) using the National Health and Nutrition Examination Survey (NHANES) III linked with Center for Medicare & Medicaid Services (CMS) data.

MATERIALS AND METHODS

Database: NHANES-CMS

The NHANES, sponsored by the National Center for Health Statistics (NCHS), consists of cross-sectional surveys providing nationally representative data on U.S. population health and nutritional status. Sampling follows a stratified, multistage probability cluster design. It includes in-home interviews for basic health and demographic information [21]. This was a retrospective cohort study whereby publicly available data was linked to restricted medical and death records and analyzed at the Research Data Center (RDC). CMS-Medicare and NDI linkage methodology are provided in Supplementary Material 1.

The present study was approved for ethical treatment of participants by the Institutional Review Board of the National Institute on Aging, Intramural Research Program.

Study sample

A participant flowchart is presented in Fig. 1, including the sample at risk and number of events. First, we included NHANES III participants aged 45+years with complete data on at least one of 19 periodontal pathogens Immunoglobulin G (IgG) humoral immune response (1988–1994, surplus serum, SPSDEPPX), mortality status and CMS-linkage data. Among 33,199 participants (aged 1–90 years) recruited in NHANES III (1988–1994) with complete socio-demographics (i.e., age and

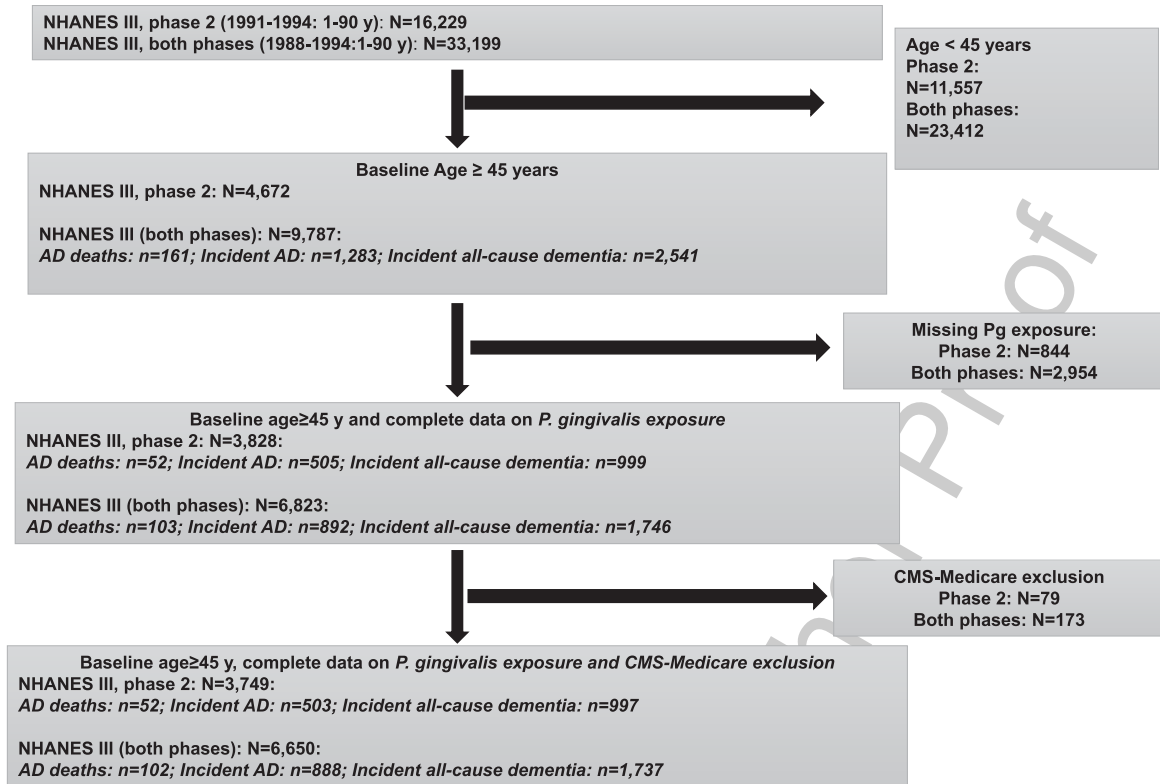


Fig. 1. Participant Flowchart. Both phases: 1988–1994; Phase 2:1991–1994. AD, Alzheimer’s disease; CMS, Centers for Medicare and Medicaid Services; NHANES III, Third National Health and Nutrition Examination Surveys.

sex), 9,787 were aged 45+years, of whom 6,823 had complete surplus serum periodontal pathogen data.

Second, 4,672 participants aged 45+years had data on humoral immune response (*IgG*) against two periodontal pathogens only for phase 2 (1991–1994) of NHANES III (DEPP file), namely *Pg* and *Aa*. Of this sub-sample, 3,828 had complete data in the DEPP file (specifically on *Pg*). Sample sizes varied for both samples, depending on exposure of interest.

Participants without CMS-Medicare data were assumed to have no event of interest until end of 2013 or censored upon death. Thus, unweighted samples consisted of 6,823 participants with *Pg* measured at both phases and 3,828 with *P. gingivalis* measured at phase 2. After exclusion due to Health Maintenance Organization (HMO) utilization, those samples were reduced to 6,650 and 3,749, respectively. Third, for PPD/AL exposures, the sample size was reduced to 5,088 (45+, both phases combined, CMS-Medicare exclusion) and PPD/AL exposures with complete both phase *P. gingivalis* and CMS-Medicare exclusion amounted to N = 4,465.

Dementia and AD onset

The CMS Chronic Condition Data Warehouse Categories included a summary file with 21 chronic conditions and varying reference time periods, numbers and types of claims to qualify, exclusions and a set of International Classification of Diseases, version 9 (ICD-9)/CPT4/HCPCS codes. AD was diagnosed using ICD-9 code 331.0 (any diagnosis on the claim) from inpatient, Skilled Nursing Patient [SNP], Home Health Agency [HHA], Health Options Program [HOP] or Carrier claims during a 3-year period. All-cause dementia was assessed using similar criteria with the following diagnostic codes: 331.0, 331.1, 331.11, 331.19, 331.2, 331.7, 290.0, 290.10, 290.11, 290.12, 290.13, 290.20, 290.21, 290.3, 290.40, 290.41, 290.42, 290.43, 294.0, 294.1, 294.10, 294.11, 294.8, and 797. We computed time-to-event starting from Medical Examination Center (MEC) examination date, using the earliest occurrence date. The summary file was available for 1999–2013 follow-up period. Using the same algorithm, we estimated AD/dementia’s earliest diagnosis

179 date during 1991–1998 [22]. Follow-up time was
180 truncated to January 1, 2014 and was expressed in
181 months.

182 Mortality from AD

183 AD mortality was a primary outcome and was
184 determined using underlying cause of death ICD-10
185 code G30 [23]. Additional AD cases were included
186 when earliest AD diagnosis date was unavailable but
187 AD-related death was assigned. Follow-up was cen-
188 sored at death or, if participants were alive by end of
189 follow-up, January 1, 2014.

190 Dental examination and clinical periodontal 191 markers

192 Attachment loss (AL) and probing pocket depth
193 (PPD) defined Pd in our study [24]. Briefly, dental
194 examiners assessed oral health during both phases of
195 MEC examinations in NHANES III. AL and PPD
196 were measured at mid-buccal and mesio-buccal sites
197 on every tooth in two randomly selected quadrants-
198 the maxilla and the mandible (Range: 14–28 teeth)
199 [25]. AL was defined as distance between cemento-
200 enamel junction and base of periodontal pocket, while
201 PPD was distance from base of periodontal pocket to
202 free gingival margin [26]. Mean AL and PPD were
203 calculated for 28 sites. Two non-missing sites per
204 tooth were required for AL/PPD measure and at least
205 one tooth measurement was required to estimate the
206 mean [26].

207 Periodontal pathogens

208 Serum immunoglobulin G (IgG) titers were mea-
209 sured for humoral immune response against 19
210 periodontal bacteria using a series of 1:1,000 serum
211 dilutions and checkerboard immunoblotting as pre-
212 viously described [27]. Those pathogens are: 1)
213 *A. Actinomycetemcomitans* (American Type Culture
214 Collection [ATCC] strains 43718, 29523, and 33384);
215 2) *P. gingivalis* (ATCC strains 3327 and 53978); 3) *T.*
216 *forsythia* (ATCC strain 43037); 4) *T. denticola* (Oral
217 Microbiology, Gothenburg, Sweden [OMGS] strain
218 3271); 5) *Campylobacter rectus* (*C. rectus*, ATCC
219 strain 33238); 6) *Eubacterium nodatum* (*E. noda-*
220 *tum*, ATCC strain 33099); 7) *Prevotella intermedia*
221 (*P. intermedia*, ATCC strain 25611); 8) *Prevotella*
222 *nigrescens* (*P. nigrescens*, ATCC strain 33563);
223 9) *Prevotella melaninogenica* (*P. melaninogenica*,
224 ATCC strain 25845); 10) *Fusobacterium nucleatum*

(*F. nucleatum*, ATCC strain 33563); 11) *Parvimonas*
225 *micra* aka *Micromonas micros* (*M. micros*, ATCC
226 strain 10953); 12) *Selenomonas noxia* (*S. noxia*,
227 ATCC strain 43541); 13) *Eikenella corrodens* (*E. cor-*
228 *rodens*, ATCC strain 23834); 14) *Capnocytophaga*
229 *ochracea* (*C. ochracea*, ATCC strain 33624); 15)
230 *Streptococcus intermedius* (*S. intermedius*, ATCC
231 strain 35037); 16) *Streptococcus oralis* (*S. oralis*,
232 ATCC strain 35037); 17) *Streptococcus mutans* (*S.*
233 *mutans*, ATCC strain 25175); 18) *Vellonella Parvula*
234 (*V. parvula*, ATCC strain 10790); 19) *Actinomyces*
235 *naeslundii* (*A. naeslundii*, ATCC strain 49340) [28].
236 IgG titers were quantified using chemiluminescent
237 signal-measuring instrument and compared to human
238 IgG standard curve [27]. Specifically, 8,153 stored
239 serums among NHANES III participants aged 40
240 years or older were analyzed at Columbia Univer-
241 sity College of Dental Medicine, New York, NY
242 between 2003 and 2006 (Phase 2 then Phase 1
243 samples). A factor analysis was conducted among
244 individuals >45 years of age, to extract indepen-
245 dent common factors, using eigenvalue and scree
246 plot criteria and varimax rotation (Supplementary
247 Material 2). These factors were entered simultane-
248 ously into models examining associations between
249 periodontal pathogens and AD mortality, AD inci-
250 dence and all-cause dementia incidence. Finally,
251 we created modified mutually-exclusive color-coded
252 clusters that were determined using cluster analy-
253 sis in a previous analysis of all available periodontal
254 pathogen data (k = 19, 40+years, NHANES III, both
255 phases) [28], and analysis also used by others [29].
256 Those clusters were defined based on correlated Log_e
257 transformed pathogen IgG titers as shown in Supple-
258 mentary Figure 1 [28]. In our present study, we first
259 summed Log_e transformed IgG to form the clusters of
260 correlated pathogen titers. Those summation clusters
261 were then z-scored to allow for better interpretation
262 in our main models.
263

264 In phase 2 of NHANES III, 9,371 surplus sera
265 on two periodontal pathogens were analyzed (*P.*
266 *gingivalis* and *A. actinomycetemcomitans*) among
267 participants aged 12 years or older of whom 3,828
268 were 45+years. Antibody concentrations were mea-
269 sured in ELISA units of IgG (EU) and were examined
270 in both untransformed and Log_e transformed metrics,
271 for comparative purposes.

272 Covariates

273 Models were stratified by baseline age group
274 (45+, 55+, and 65+) and sex. Demographic,

socio-economic, social support, lifestyle and health-related factors, dietary quality and nutritional biomarkers were potential confounders included in all models (See Supplementary Material 2 for details).

Statistical analysis

We performed analyses using Stata 15.0 (Stata Corp, College Station, TX) [30]. We accounted for survey design complexity by incorporating 6-year (NHANES III, 1988–1994) and 3-year (NHANES III, 1991–1994) primary sampling units and strata. Standard errors were estimated using Taylor series linearization (i.e., *svy*: commands) [30]. Multivariate imputed data ($m = 5$ imputations, 10 iterations) with chained equations [31] estimated means and proportions across age groups and measures of association, after adjusting for sampling design complexity.

The main exposures of interest were 19 periodontal pathogens added simultaneously (Model 1) and separately (Model 2) for the entire NHANES III sample (1988–1994). Factors extracted from those 19 periodontal pathogens, as well as pre-determined color-coded clusters, were also considered as predictors of interest. For phase 2 NHANES III, *A. Actinomycetemcomitans* and *P. gingivalis* were main predictors, analyzed as standardized z-scores (as is and Log_e transformed). These periodontal pathogens were then entered as predictors (both phases and phase 2, separately) into linear regression models with periodontitis measured with AL and PPD as outcomes while adjusting for all covariates. Finally, means of AL/PPD were considered as separate predictors in main causal models, specifically incident AD and all-cause dementia due to smaller sample size for AD mortality outcome. In those models, defining time-to-event from any age ≥ 45 years since baseline visit (i.e., delayed entry) until death or censoring or outcome of interest (AD death, AD incidence, all-cause dementia incidence), we conducted Cox proportional hazards models for three outcomes stratifying separately by sex and baseline age to ≥ 45 , ≥ 55 , and ≥ 65 years. We present fully adjusted models accounting for demographic, socio-economic, lifestyle/social support factors, nutritional biomarkers, and health-related factors (Supplementary Material 2). Weighted mean times of follow-up are estimated from weighted person-months of follow-up and the weighted sample in each model ([Person-

months_(weighted)]/[Persons_(weighted)]). A type I error of 0.05 was considered for statistical significance and 0.10 for borderline (or marginal) significance. Multiple testing adjustment was done using a familywise Bonferroni approach while accounting for outcome multiplicity (e.g., AL/PPD: 2 outcomes; incident AD/all-cause dementia/AD mortality: 3 outcomes) and assuming that each exposure, model and strata was a distinctive hypothesis [32].

RESULTS

Participant characteristics by age group and sex

Cumulative incidence (weighted) of all three outcomes increased linearly with baseline age, with AD dementia, all-cause dementia and AD mortality reaching 18%, 38%, and 3% in the 65+ baseline age group, respectively. Women in this sample were more likely to be older, and baseline age was also directly related to Non-Hispanic white race, smaller household size, higher proportion widowed, larger means of AL and PPD, and higher proportions completely or partially edentulous (Table 1 and Supplementary Table 1). However, most periodontal pathogen titers were either unrelated or inversely linked to age, with the clearest inverse relationship shown for Orange Blue and Yellow Orange clusters, and for *Factor 3* comprised of *E. nodatum* and *A. naeslundii* IgG titers. Moreover, socio-economic status was associated with younger age, while age was directly related to better dietary quality as measured by the 1995 Healthy Eating Index, co-morbidity and AL, reduced physical activity, smoking and drug use, lower prevalence of obesity, reduced mean of 25(OH)D coupled with increased levels of folate, vitamin A, vitamin E, total carotenoids, and ferritin.

Periodontal pathogens' association with AD mortality, AD and all-cause dementia incidence

After correction for multiple testing (Table 2), Phase 2 *P. gingivalis* IgG titers (un-transformed, z-scores) were associated with increased risk for incident AD dementia, particularly among women (1 SD = 212, HR = 1.14, 95% CI: 1.05–1.23, $p = 0.004$) and individuals above 55 (HR = 1.06) or 65 years (HR = 1.12) at baseline. Log_e transformed *P. gingivalis* IgG titers were marginally associated with increased risk for AD mortality in 65+ age group, while the reverse was true for *A. Actinomycetemcomi-*

Table 1

Baseline characteristics of selected participants by age group, NHANES III, 1988–1994 [N = 3,749 (phase 2:1991–1994); N = 6,650 (both phases)]^a

Selected participant characteristics	Age group (y)			<i>p</i> -value*	
	45–55	55–65	65+	(Design-based F-test) ^b	
				55–65 versus 45–55	65+ versus 45–55
Unweighted N (both phases)	(N = 1,701) 25.6%	(N = 1,698) 25.5%	(N = 3,251) 48.9%		
<i>Cumulative incidence of AD and all-cause dementia and of AD mortality, weighted %</i>					
<i>AD dementia</i>	1.7 ± 0.3	11.2 ± 1.1	18.3 ± 0.9	<0.001	<0.001
<i>All-cause dementia</i>	4.3 ± 0.5	18.9 ± 1.3	37.5 ± 1.1	<0.001	<0.001
<i>AD mortality</i>	0.1 ± 0.1	1.1 ± 0.4	2.9 ± 0.4	0.028	0.001
<i>Dental measures</i>					
Periodontitis, mean ± SE	(N = 1,435)	(N = 1,225)	(N = 1,805)		
Attachment Loss	1.45 ± 0.05	1.76 ± 0.07	1.98 ± 0.07	0.03	<0.001
Probing Pocket Depth	1.50 ± 0.03	1.53 ± 0.04	1.45 ± 0.03	0.29	0.28
Factors (z-scores), mean ± SE	(N = 1,597)	(N = 1,604)	(N = 3,077)		
Factor 1	-0.05 ± 0.09	+0.00 ± 0.09	0.02 ± 0.08	0.35	0.24
Factor 2	-0.05 ± 0.04	-0.09 ± 0.04	-0.15 ± 0.04	0.46	0.063
Factor 3	+0.21 ± 0.04	+0.13 ± 0.04	-0.04 ± 0.03	0.10	<0.001
Factor 4	-0.07 ± 0.06	-0.15 ± 0.04	-0.09 ± 0.04	0.09	0.62
Factor 5	+0.05 ± 0.07	-0.04 ± 0.05	+0.03 ± 0.04	0.12	0.84
Clusters (z-scores), mean ± SE	(N = 1,655)	(N = 1,655)	(N = 3,169)		
Orange Red	-0.097 ± 0.04	-0.150 ± 0.040	-0.183 ± 0.036	0.35	0.078
Red Green	-0.031 ± 0.010	-0.071 ± 0.085	-0.046 ± 0.071	0.55	0.83
Yellow Orange	+0.02 ± 0.06	-0.02 ± 0.06	-0.10 ± 0.05	0.51	0.020
Orange Blue	+0.19 ± 0.04	+0.10 ± 0.05	-0.10 ± 0.04	0.08	<0.001
Dentate status	(N = 1,701)	(N = 1,698)	(N = 3,251)		
Completely edentulous	9.2 ± 1.11	19.0 ± 1.24	32.2 ± 1.87	---	---
Edentulous in one arch	10.5 ± 1.22	14.5 ± 1.07	14.8 ± 0.95	<0.001	<0.001
Teeth complete	80.3 ± 1.48	66.5 ± 1.54	52.9 ± 2.04	0.001	<0.001
<i>Periodontal pathogen IgG</i>					
Phase 2	(N = 895)	(N = 914)	N = 1,826)		
<i>P. gingivalis</i>					
Continuous	109.2 ± 6.34	105.2 ± 4.2	122.3 ± 7.55	0.49	0.16
Log _e transformed	4.49 ± 0.03	4.47 ± 0.02	4.51 ± 0.02	0.30	0.64
<i>A. Actinomycetemcomitans (Aa)</i>					
Continuous	99.3 ± 5.53	102.8 ± 5.15	94.5 ± 2.87	0.66	0.39
Log _e transformed	4.45 ± 0.03	4.46 ± 0.02	4.44 ± 0.02	0.79	0.61
<i>Both Phases, Log_e transformed^c</i>					
<i>P. Gingivalis</i> (Pg) mix	(N = 1,622–1,692)	(N = 1,617–1,685)	(N = 3,116–3,227)		
<i>P. Intermedia</i> (Pi)	5.7 ± 0.1	5.6 ± 0.1	5.6 ± 0.1	0.44	0.34
<i>P. Nigrescens</i> (Pn)	5.6 ± 0.1	5.5 ± 0.1	5.3 ± 0.0	0.57	0.002
<i>T. Forsythia</i> (Tf)	5.3 ± 0.1	5.2 ± 0.1	5.2 ± 0.1	0.20	0.06
<i>A. Actinomycetemcomitans</i> (Aa) mix	4.8 ± 0.1	4.7 ± 0.1	4.7 ± 0.1	0.37	0.76
<i>F. Nucleatum</i> (Fn)	6.7 ± 0.1	6.7 ± 0.1	6.6 ± 0.1	0.65	0.41
<i>S. Oralis</i> (So)	4.4 ± 0.1	4.4 ± 0.1	4.4 ± 0.1	0.78	0.15
<i>M. Micros</i> (Mm)	4.3 ± 0.1	4.3 ± 0.1	4.2 ± 0.1	0.64	0.54
<i>C. Rectus</i> (Cr)	5.2 ± 0.1	5.2 ± 0.1	5.1 ± 0.1	0.52	0.13
<i>E. Corrodens</i> (Ec)	4.4 ± 0.1	4.3 ± 0.1	4.4 ± 0.1	0.11	0.73
<i>E. Nodatum</i> (En)	5.2 ± 0.1	5.2 ± 0.1	5.3 ± 0.1	0.63	0.15
<i>S. Intermedius</i> (Si)	7.3 ± 0.1	7.1 ± 0.1	6.8 ± 0.1	0.09	<0.001
<i>C. Ochracea</i> (Co)	5.2 ± 0.1	5.1 ± 0.1	5.0 ± 0.1	0.68	0.002
<i>V. Parvula</i> (Vp)	5.0 ± 0.1	4.8 ± 0.1	4.7 ± 0.0	0.052	0.002
<i>A. Naeslundii</i> (An)	3.6 ± 0.1	3.7 ± 0.1	3.8 ± 0.1	0.97	0.021
<i>P. Melaninogenica</i> (Pm)	6.1 ± 0.1	5.9 ± 0.1	5.7 ± 0.1	0.10	<0.001
<i>S. Noxia</i> (Sn)	5.3 ± 0.1	5.4 ± 0.1	5.4 ± 0.1	0.99	0.35
<i>T. Denticola</i> (Td)	3.7 ± 0.2	3.6 ± 0.1	3.5 ± 0.1	0.21	0.21
<i>S. Mutans</i> (Sm)	4.9 ± 0.1	4.8 ± 0.1	4.8 ± 0.1	0.35	0.43
	4.5 ± 0.1	4.5 ± 0.1	4.4 ± 0.1	0.82	0.40
<i>Socio-demographic characteristics</i>					
Age (y)	(N = 1,701)	(N = 1,698)	(N = 3,251)		
Sex, % male	49.1 ± 0.11	59.3 ± 0.09	73.6 ± 0.24	<0.001	<0.001
	48.5 ± 1.75	44.8 ± 1.12	41.4 ± 1.12	0.07	0.002

	(N = 1,701)	(N = 1,698)	(N = 3,251)		
Race/ethnicity					
Non-Hispanic White	79.7 ± 1.80	79.0 ± 2.06	86.6 ± 1.30	--	--
Non-Hispanic Black	8.8 ± 0.74	9.6 ± 0.90	7.3 ± 0.76	0.39	0.04
Mexican-American	4.3 ± 0.43	2.9 ± 0.31	1.9 ± 0.17	0.003	<0.001
Other	7.3 ± 1.46	8.5 ± 1.97	4.3 ± 0.85	0.54	0.02
Urban/rural area of residence	(N = 1,701)	(N = 1,698)	(N = 3,251)		
Urban	50.1 ± 4.69	46.1 ± 4.53	43.2 ± 5.11	0.18	0.06
Rural	49.1 ± 4.69	53.9 ± 4.53	56.8 ± 5.11	--	--
Household size	2.9 ± 0.06	2.5 ± 0.04	1.9 ± 0.03	<0.001	<0.001
Marital status	(N = 1,701)	(N = 1,698)	(N = 3,251)		
Never married	5.4 ± 0.97	3.1 ± 0.43	4.0 ± 0.41	0.02	0.93
Married	73.7 ± 2.12	71.8 ± 1.5	54.9 ± 1.45	--	--
Divorced	13.7 ± 1.72	10.4 ± 1.06	5.4 ± 0.57	0.14	0.003
Widowed	2.3 ± 0.39	9.7 ± 0.9	33.3 ± 1.17	<0.001	<0.001
Other	4.8 ± 0.61	4.9 ± 0.85	2.5 ± 0.39	0.82	0.08

25(OH)D, 25-hydroxyvitamin D; AD, Alzheimer's disease; EU, ELISA units; HEI, Healthy Eating Index; HS, high school; IgG, Immunoglobulin G; MAR, Mean Adequacy Ratio; NHANES, National Health and Nutrition Examination Surveys. ^a Values are weighted means ± SEM or percent ± SEP, taking into account sampling design complexity (PSU and strata), across 5 imputations with 10 iterations. ^b Design-based F-test accounting for design complexity in terms of sampling weights, PSU and stratum. For categorical variables, this was the equivalent of a χ^2 test of independence restricting the sample first to 55–64/45–54, then to 65+/45–54. For continuous variables, it was the equivalent of a Wald test in a linear regression model with the variable being the outcome predicted by age group and in which 45–54 years was the referent category to which “55–64” and “65+” were compared. ^c SD of Log_e transformed periodontal pathogens across groups ranged between 1.2–1.3 (*Co*, *Vp*) and 1.8–2.0 (*Pg*, *En*, *An*), with the remaining ranging between 1.4 and 1.6.

370 *tans* IgG (both transformed and untransformed, 65+).
 371 When examining all 19 periodontal pathogens (Log_e
 372 transformed, z-scored, 1988–1994) in relation to the
 373 three dementia outcomes of interest (Supplementary
 374 Tables 2 and 3), and upon multiple testing adjust-
 375 ment, we found that *S. oralis* IgG was linked with
 376 increased risk for all-cause dementia among men,
 377 a pattern observed among women for *E. corrodens*
 378 IgG (Model 1: all pathogens entered). Similarly, *C.*
 379 *rectus* IgG was associated with increased risk for
 380 all-cause dementia in all age groups, a pattern that
 381 was consistent between models 1 and 2 among the
 382 older group (65+). *C. rectus* IgG was also marginally
 383 and directly associated with incident AD risk in the
 384 55+ age group (Model 1), while *S. intermedius* was
 385 marginally and inversely associated with incident
 386 AD risk among women (Model 1). For AD mortal-
 387 ity (Table 3), most of our findings emerged when
 388 each periodontal pathogen was entered separately
 389 into the model (Model 2). Most notably, *P. gingivalis*
 390 IgG (Log_e transformed, z-score) was associated with
 391 increased AD mortality risk among those aged 65+ at
 392 baseline (1 SD = 2.03, HR = 1.36, 95% CI: 1.10–1.69,
 393 *p* = 0.010), as was the case for *P. melaninogenica*
 394 IgG (1 SD = 1.28, HR = 1.43, 95% CI: 1.11–1.85,
 395 *p* = 0.009). The latter finding was further strength-
 396 ened by adding the remaining 18 titers into the
 397 model (HR = 1.73, *p* = 0.005). Consistent with all-
 398 cause dementia (Supplementary Table 2), Table 3 also
 399 indicates that *So* was directly related to AD mortality

400 risk among men (Model 2). Additionally, AD mor-
 401 tality risk was increased with higher *S. intermedius*
 402 IgG.

403 Periodontal pathogen factor and clusters and 404 their association with AD mortality, AD and 405 all-cause dementia incidence

406 Using factor analysis and pre-defined clusters
 407 (Table 4), our results indicated that AD incidence
 408 was associated with Factor 4 in the 65+ age group,
 409 which loaded highly on *C. rectus* and *P. gingivalis*
 410 titers (aHR = 1.22; 95% CI, 1.04–1.43, *p* = 0.012).
 411 In this model, the effect of 1 SD increase in Fac-
 412 tor 4 on AD incidence was equivalent to two years
 413 of aging on the Log_e(HR) scale. Moreover, AD
 414 mortality risk was increased with higher baseline Fac-
 415 tor 2 in that age group (per SD, aHR = 1.46; 95%
 416 CI, 1.09–1.96, *p* = 0.017) which loaded highly on
 417 *P. gingivalis*, *P. intermedia*, *P. nigrescens*, *F. nucla-*
 418 *tum*, *C. rectus*, *S. intermedius*, *C. ochracea* and *P.*
 419 *melaninogenica* titers. In both 55+ and 65+ age group,
 420 Orange-Red cluster (*P. melaninogenica*, *P. interme-*
 421 *dia*, *P. nigrescens*, *P. gingivalis*) was associated with
 422 increased AD mortality risk, while Red-Green cluster
 423 (*T. forsythia*, *T. denticola*, *A. actinomycetemcomi-*
 424 *tans*, *E. corrodens*, *S. noxia*, *V. parvula*, *C. rectus*) was
 425 only marginally associated with AD and all-cause
 426 dementia among women (*p* < 0.033), after correction
 427 for multiple testing.

Table 2

P. gingivalis and *A. actinomycetemcomitans* serum IgG's association with incident all-cause and Alzheimer's disease (AD) dementia and with AD mortality in multiple Cox proportional hazards model, overall and stratified by sex and race: NHANES III, 1991–1994^a

	z-scored periodontal pathogen IgG (Phase II) ^b			z-scored, Log _e transformed periodontal pathogen IgG (Phase II) ^b		
	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p
All-cause dementia^c						
Men (N = 1,646)						
<i>P. gingivalis</i>	-0.19	(0.17)	0.31	-0.12	(0.10)	0.26
<i>A. actinomycetemcomitans</i>	+0.03	(0.06)	0.62	+0.03	(0.06)	0.66
Women (N = 1,979)						
<i>P. gingivalis</i>	+0.07	(0.07)	0.34	-0.04	(0.10)	0.69
<i>A. actinomycetemcomitans</i>	+0.05	(0.05)	0.36	+0.03	(0.07)	0.62
45+ at baseline (N = 3,625)						
<i>P. gingivalis</i>	-0.01	0.11	0.92	-0.08	(0.06)	0.19
<i>A. actinomycetemcomitans</i>	+0.04	(0.04)	0.34	+0.03	(0.06)	0.60
55+ at baseline (N = 2,731)						
<i>P. gingivalis</i>	+0.01	(0.09)	0.88	-0.08	(0.06)	0.21
<i>A. actinomycetemcomitans</i>	-0.01	(0.05)	0.91	-0.01	(0.05)	0.81
65+ at baseline (N = 1,817)						
<i>P. gingivalis</i>	+0.04	(0.07)	0.59	-0.06	(0.06)	0.33
<i>A. actinomycetemcomitans</i>	+0.12	(0.06)	0.051	+0.08	(0.06)	0.19
AD dementia^d						
Men (N = 1,646)						
<i>P. gingivalis</i>	-0.05	(0.21)	0.81	-0.11	(0.12)	0.37
<i>A. actinomycetemcomitans</i>	+0.10	(0.11)	0.34	+0.11	(0.11)	0.34
Women (N = 1,979)						
<i>P. gingivalis</i>	+0.13	(0.04)	0.004**	+0.04	(0.06)	0.58
<i>A. actinomycetemcomitans</i>	-0.03	(0.11)	0.81	-0.09	(0.11)	0.40
45+ at baseline (N = 3,625)						
<i>P. gingivalis</i>	+0.06	(0.03)	0.034	-0.04	(0.06)	0.50
<i>A. actinomycetemcomitans</i>	+0.01	(0.06)	0.84	-0.00	(0.07)	0.95
55+ at baseline (N = 2,731)						
<i>P. gingivalis</i>	+0.06	(0.02)	0.015**	-0.03	(0.06)	0.64
<i>A. actinomycetemcomitans</i>	-0.04	(0.08)	0.62	-0.07	(0.07)	0.28
65+ at baseline (N = 1,817)						
<i>P. gingivalis</i>	+0.11	(0.03)	0.003**	+0.03	(0.07)	0.72
<i>A. actinomycetemcomitans</i>	+0.07	(0.07)	0.35	+0.02	(0.07)	0.73
AD mortality^e						
45+ at baseline (N = 3,625)						
<i>P. gingivalis</i>	+0.11	(0.07)	0.13	+0.14	(0.14)	0.31
<i>A. actinomycetemcomitans</i>	-0.03	(0.34)	0.94	-0.27	(0.30)	0.31
55+ at baseline (N = 2,731)						
<i>P. gingivalis</i>	+0.09	(0.06)	0.13	+0.16	(0.14)	0.27
<i>A. actinomycetemcomitans</i>	-0.02	(0.35)	0.95	-0.33	(0.36)	0.38
65+ at baseline (N = 1,817)						
<i>P. gingivalis</i>	+0.19	(0.15)	0.21	+0.35	(0.15)	0.033*
<i>A. actinomycetemcomitans</i>	-2.61	(0.73)	0.002**	-1.23	(0.28)	<0.001**

25(OH)D, 25-hydroxyvitamin D; AD, Alzheimer's disease; EU, ELISA units; HEI, Healthy Eating Index; HR, hazard ratio; HS, high school; IgG, Immunoglobulin G; MAR = Mean Adequacy Ratio; NHANES = National Health and Nutrition Examination Surveys. ^aModels were adjusted for age, sex, race/ethnicity, poverty income ratio, education (years), urban-rural area of residence, household size, marital status, nutritional factors (HEI, MAR), nutritional biomarkers (25(OH)D, folate, vitamin C, vitamin A, total carotenoids, vitamin E, ferritin, selenium and normalized calcium), lifestyle (smoking, drug use, alcohol, physical activity), health-related factors (self-rated health, comorbidity index, allostatic load, weight status), dentate status and social support variables. Covariates (other than exposures) were imputed and analysis is across 5 imputations with 10 iterations. ^bStandardized into z-scores. 1 SD of untransformed Pg is 434 (45+), 148 (55+), 276 (65+), 605 (Men), 212 (Women); 1 SD of untransformed Aa is 87 (45+), 82 (55+), 72 (65+), 85 (Men), 88 (Women). 1 SD of Log_e transformed Pg is ~0.85–0.90 for all groups; 1 SD of Log_e transformed Aa is ~0.85–0.90 for all groups. ^c997 unweighted incident dementia cases for 45+, weighted mean follow-up time: 185 months. ^d503 unweighted incident AD cases for 45+, weighted mean follow-up time: 189 months. ^e52 unweighted AD deaths for 45+, weighted mean follow-up time: 192 months. **p* < 0.033, marginally significant after correction for multiple testing; ***p* < 0.016, significant after correction for multiple testing.

Table 3

Periodontal pathogens' serum IgG association with AD mortality in multiple Cox proportional hazards model, overall and restricted by baseline age group and sex: NHANES III, 1988–1994^{a,b}

	≥45 y ^c			≥55 y ^d			≥65 y ^e			Men ^f			Women ^g		
	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p
AD mortality															
<i>P. Gingivalis</i> mix															
Model 1	+0.14	(0.13)	0.88	+0.15	(0.14)	0.28	+0.35	(0.17)	0.046	+0.25	(0.30)	0.40	+0.02	(0.25)	0.93
Model 2	+0.15	(0.10)	0.16	+0.17	(0.11)	0.13	+0.31	(0.11)	0.010**	+0.33	(0.20)	0.11	-0.02	(0.13)	0.87
<i>P. Intermedia</i>															
Model 1	-0.08	(0.25)	0.74	-0.14	(0.27)	0.61	-0.19	(0.32)	0.57	+0.11	(0.52)	0.84	-0.04	(0.38)	0.93
Model 2	+0.20	(0.14)	0.15	+0.19	(0.13)	0.16	+0.29	(0.13)	0.026*	+0.02	(0.14)	0.86	+0.11	(0.19)	0.58
<i>P. Nigrescens</i>															
Model 1	+0.18	(0.20)	0.36	+0.28	(0.21)	0.20	+0.18	(0.29)	0.54	-0.10	(0.66)	0.87	+0.10	(0.26)	0.69
Model 2	+0.22	(0.13)	0.085	+0.25	0.13	0.063	+0.33	(0.14)	0.023*	+0.43	(0.21)	0.049	+0.11	(0.11)	0.46
<i>T. Forsythia</i>															
Model 1	+0.43	(0.22)	0.058	+0.41	(0.21)	0.060	+0.39	(0.25)	0.13	+0.86	(0.57)	0.14	+0.64	(0.26)	0.019*
Model 2	+0.23	(0.14)	0.10	+0.22	(0.14)	0.12	+0.34	(0.16)	0.036	+0.49	(0.22)	0.031*	+0.13	(0.17)	0.43
<i>A. Actinomycetemcomitans (Aa)</i> mix															
Model 1	-0.38	(0.26)	0.15	-0.48	(0.27)	0.078	-0.71	(0.45)	0.12	-0.05	(0.35)	0.88	-0.80	(0.38)	0.039
Model 2	-0.03	(0.15)	0.82	-0.10	(0.15)	0.52	-0.02	(0.19)	0.92	+0.59	(0.28)	0.042	-0.31	(0.20)	0.12
<i>F. Nucleatum</i>															
Model 1	-0.08	(0.28)	0.77	-0.04	(0.28)	0.89	+0.26	(0.35)	0.46	-0.77	(0.48)	0.12	+0.17	(0.40)	0.68
Model 2	+0.03	(0.13)	0.84	+0.02	(0.13)	0.86	+0.18	(0.14)	0.18	+0.38	(0.19)	0.053	-0.04	(0.16)	0.78
<i>S. Oralis</i>															
Model 1	+0.10	(0.26)	0.69	+0.03	(0.26)	0.90	+0.14	(0.31)	0.66	+1.14	(0.46)	0.019*	-0.29	(0.29)	0.32
Model 2	+0.05	(0.13)	0.72	+0.02	(0.13)	0.87	+0.16	(0.13)	0.22	+0.64	(0.25)	0.014**	-0.16	(0.16)	0.31
<i>M. Micros</i>															
Model 1	+0.21	(0.19)	0.29	+0.17	(0.20)	0.41	-0.20	(0.24)	0.39	-0.34	(0.32)	0.29	+0.42	(0.28)	0.15
Model 2	+0.15	(0.10)	0.16	+0.10	(0.12)	0.42	+0.07	(0.14)	0.64	+0.19	(0.21)	0.38	+0.13	(0.14)	0.35
<i>C. Rectus</i>															
Model 1	-0.13	(0.18)	0.49	-0.03	(0.19)	0.89	-0.11	(0.26)	0.69	-0.80	(0.52)	0.13	+0.02	(0.22)	0.91
Model 2	+0.07	(0.12)	0.57	+0.01	(0.16)	0.96	+0.23	(0.14)	0.10	+0.18	(0.22)	0.42	-0.04	(0.14)	0.76
<i>E. Corrodens</i>															
Model 1	+0.12	(0.30)	0.68	-0.01	(0.28)	0.98	+0.27	(0.35)	0.44	-0.13	(0.39)	0.74	+0.13	(0.38)	0.74
Model 2	+0.08	(0.18)	0.67	+0.02	(0.17)	0.90	+0.19	(0.15)	0.23	+0.29	(0.24)	0.23	-0.02	(0.24)	0.92
<i>E. Nodatum</i>															
Model 1	-0.01	(0.19)	0.96	-0.01	(0.21)	0.98	+0.15	(0.22)	0.50	-0.51	(0.40)	0.21	-0.12	(0.31)	0.71
Model 2	+0.06	(0.17)	0.73	+0.02	(0.17)	0.90	+0.20	(0.19)	0.31	+0.32	(0.19)	0.094	-0.12	(0.25)	0.64

(Continued)

Table 3
(Continued)

	≥45 y ^c			≥55 y ^d			≥65 y ^e			Men ^f			Women ^g		
	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p
<i>S. Intermedius</i>															
Model 1	-0.09	(0.24)	0.71	-0.23	(0.23)	0.33	-0.11	(0.31)	0.72	+0.93	(0.37)	0.78	-0.35	(0.32)	0.28
Model 2	+0.04	(0.12)	0.72	-0.02	(0.12)	0.88	+0.15	(0.15)	0.33	+0.72	(0.21)	0.001**	-0.19	(0.16)	0.26
<i>C. Ochracea</i>															
Model 1	+0.08	(0.17)	0.64	+0.10	(0.18)	0.56	+0.07	(0.23)	0.76	+0.10	(0.34)	0.78	-0.04	(0.22)	0.86
Model 2	+0.05	(0.13)	0.72	+0.06	(0.14)	0.68	+0.09	(0.17)	0.58	+0.21	(0.16)	0.19	-0.08	(0.17)	0.61
<i>V. Parvula</i>															
Model 1	-0.13	(0.28)	0.65	+0.02	(0.25)	0.93	-0.10	(0.29)	0.73	+0.08	(0.41)	0.85	-0.06	(0.32)	0.85
Model 2	+0.06	(0.14)	0.68	+0.02	(0.16)	0.91	+0.17	(0.15)	0.26	+0.46	(0.23)	0.051	-0.10	(0.18)	0.58
<i>A. Naeslundii</i>															
Model 1	+0.03	(0.18)	0.89	-0.02	(0.19)	0.92	-0.04	(0.23)	0.86	+0.57	(0.51)	0.27	-0.06	(0.21)	0.77
Model 2	+0.04	(0.16)	0.80	+0.22	(0.14)	0.12	+0.11	(0.19)	0.57	+0.48	(0.20)	0.020*	-0.18	(0.21)	0.39
<i>P. Melaninogenica</i>															
Model 1	+0.30	(0.21)	0.17	+0.36	(0.22)	0.11	+0.55	(0.19)	0.005**	+0.20	(0.38)	0.59	+0.42	(0.27)	0.12
Model 2	+0.20	(0.13)	0.14	+0.22	(0.14)	0.12	+0.36	(0.13)	0.009**	+0.41	(0.20)	0.047	+0.15	(0.18)	0.40
<i>S. Noxia</i>															
Model 1	-0.15	(0.22)	0.50	-0.14	(0.22)	0.53	-0.14	(0.25)	0.57	-0.04	(0.32)	0.91	-0.34	(0.31)	0.28
Model 2	+0.02	(0.12)	0.87	+0.04	(0.13)	0.77	+0.13	(0.13)	0.31	+0.37	(0.21)	0.092	-0.16	(0.17)	0.36
<i>T. Denticola</i>															
Model 1	-0.10	(0.25)	0.68	-0.06	(0.24)	0.79	-0.18	(0.25)	0.48	-0.15	(0.32)	0.64	-0.03	(0.32)	0.92
Model 2	-0.09	(0.13)	0.49	-0.09	(0.13)	0.49	-0.05	(0.13)	0.70	+0.17	(0.25)	0.50	-0.13	(0.18)	0.48
<i>S. Mutans</i>															
Model 1	-0.26	(0.23)	0.27	-0.26	(0.24)	0.30	-0.19	(0.27)	0.49	-0.40	(0.29)	0.18	+0.03	(0.27)	0.90
Model 2	-0.07	(0.13)	0.59	-0.08	(0.13)	0.52	+0.02	(0.14)	0.86	+0.35	(0.19)	0.074	-0.22	(0.16)	0.17

AD, Alzheimer's disease; BMI, body mass index; HR, hazard ratio; HS, high school; IgG, Immunoglobulin G; NHANES, National Health and Nutrition Examination Surveys. ^aModels were adjusted for age, sex, race/ethnicity, poverty income ratio, education (years), urban-rural area of residence, household size, marital status, nutritional factors (HEI, MAR), nutritional biomarkers (25(OH)D, folate, vitamin C, vitamin A, total carotenoids, vitamin E, ferritin, selenium and normalized calcium), lifestyle (smoking, drug use, alcohol, physical activity), health-related factors (self-rated health, co-morbidity index, allostatic load, weight status), dentate status and social support variables, as well as Phase of NHANES III. Covariates (other than exposures) were imputed and analysis is across 5 imputations with 10 iterations. Model 1: adjusted for all other periodontal pathogens; Model 2: one periodontal pathogen at a time. ^bPeriodontal pathogen exposures were Log_e transformed and then standardized into z-scores. ^cUnweighted N = 6,277–6,581, weighted mean follow-up time: 200 months; ^dUnweighted N = 4,681–4,912, weighted mean follow-up time = 177 months; ^e Unweighted N = 3,077–3,229, weighted mean follow-up time: 148 months; ^fUnweighted N = 2,924–3,088, weighted mean follow-up time: 196 months ^gUnweighted N = 3,353–3,517, weighted mean follow-up time: 205 months. **p* < 0.033, marginally significant after correction for multiple testing; ***p* < 0.016, significant after correction for multiple testing.

Table 4

Periodontal pathogens' serum IgG (Factor scores and pre-defined clusters) association with AD mortality, AD incidence and all-cause dementia incidence in multiple Cox proportional hazards model, overall and restricted by baseline age group and sex: NHANES III, 1988–1994^{a,b}

	≥ 45 y ^c			≥ 55 y ^d			≥ 65 y ^e			Men ^f			Women ^g		
	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p
Factor scores															
AD mortality															
Factor 1: So/Sm/Ec	-0.12	(0.16)	0.48	-0.23	(0.15)	0.14	-0.08	(0.17)	0.61	+0.56	(0.40)	0.17	-0.30	(0.24)	0.24
Factor 2: Pi/Pn/Pm	+0.26	(0.14)	0.073	+0.31	(0.15)	0.043	+0.38	(0.15)	0.017**	+0.43	(0.29)	0.15	+0.19	(0.20)	0.35
Factor 3: An/En	+0.06	(0.19)	0.75	+0.02	(0.19)	0.90	+0.13	(0.23)	0.58	+0.44	(0.24)	0.081	-0.14	(0.27)	0.35
Factor 4: Pg/Cr	+0.12	(0.14)	0.40	+0.16	(0.15)	0.31	+0.19	(0.16)	0.24	-0.29	(0.28)	0.30	+0.16	(0.16)	0.32
Factor 5: Co/Sn	+0.04	(0.18)	0.82	+0.23	(0.13)	0.081	+0.18	(0.19)	0.34	-0.44	(0.39)	0.27	+0.17	(0.24)	0.48
AD incidence															
Factor 1: So/Sm/Ec	+0.06	(0.06)	0.34	+0.00	(0.07)	0.97	+0.06	(0.08)	0.48	+0.08	(0.14)	0.57	+0.08	(0.07)	0.25
Factor 2: Pi/Pn/Pm	+0.01	(0.05)	0.92	+0.01	(0.05)	0.86	+0.02	(0.06)	0.80	-0.00	(0.13)	1.00	-0.03	(0.07)	0.25
Factor 3: An/En	-0.08	(0.06)	0.20	-0.10	(0.06)	0.14	-0.11	(0.07)	0.13	-0.06	(0.11)	0.59	-0.10	(0.08)	0.25
Factor 4: Pg/Cr	+0.10	(0.06)	0.14	+0.11	(0.07)	0.11	+0.20	(0.08)	0.012**	+0.12	(0.15)	0.44	+0.17	(0.08)	0.038
Factor 5: Co/Sn	-0.06	(0.09)	0.49	-0.02	(0.10)	0.83	-0.02	(0.12)	0.89	-0.05	(0.14)	0.72	-0.06	(0.09)	0.52
All-cause dementia incidence															
Factor 1: So/Sm/Ec	+0.10	(0.05)	0.062	+0.06	(0.05)	0.28	+0.09	(0.05)	0.073	+0.11	(0.09)	0.22	+0.10	(0.06)	0.073
Factor 2: Pi/Pn/Pm	-0.05	(0.04)	0.23	-0.03	(0.04)	0.55	-0.02	(0.04)	0.70	+0.02	(0.08)	0.83	-0.10	(0.06)	0.070
Factor 3: An/En	-0.07	(0.04)	0.15	-0.06	(0.04)	0.19	-0.07	(0.05)	0.16	-0.04	(0.08)	0.66	-0.08	(0.06)	0.17
Factor 4: Pg/Cr	+0.02	(0.05)	0.67	+0.04	(0.05)	0.39	+0.09	(0.06)	0.12	-0.10	(0.11)	0.33	+0.06	(0.06)	0.32
Factor 5: Co/Sn	-0.09	(0.06)	0.13	-0.06	(0.06)	0.39	-0.05	(0.07)	0.48	-0.15	(0.09)	0.10	-0.02	(0.07)	0.72
Pre-defined clusters															
AD mortality															
Cluster 1: Orange-Red	+0.36	(0.16)	0.031*	+0.44	(0.18)	0.016**	+0.56	(0.17)	0.002**	+0.46	(0.47)	0.34	+0.29	(0.27)	0.28
Cluster 2: Red-Green	-0.22	(0.30)	0.47	-0.20	(0.31)	0.54	-0.16	(0.32)	0.61	-0.41	(0.75)	0.59	-0.18	(0.35)	0.61
Cluster 3: Yellow-Orange	+0.01	(0.20)	0.97	-0.07	(0.21)	0.73	-0.09	(0.25)	0.72	+0.38	(0.42)	0.37	-0.06	(0.23)	0.81
Cluster 4: Orange-Blue	+0.05	(0.18)	0.79	+0.01	(0.18)	0.94	+0.11	(0.22)	0.63	+0.34	(0.24)	0.16	-0.15	(0.26)	0.55

(Continued)

Table 4
(Continued)

	≥45 y ^c			≥55 y ^d			≥65 y ^e			Men ^f			Women ^g		
	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p	Log _e (HR)	(SE)	p
AD incidence															
Cluster 1: Orange-Red	+0.02	(0.07)	0.75	+0.05	(0.08)	0.52	+0.05	(0.09)	0.57	-0.04	(0.14)	0.79	-0.00	(0.09)	0.99
Cluster 2: Red-Green	+0.16	(0.10)	0.12	+0.14	(0.09)	0.12	+0.22	(0.11)	0.052	-0.17	(0.21)	0.40	+0.30	(0.13)	0.028*
Cluster 3: Yellow-Orange	-0.12	(0.09)	0.21	-0.16	(0.09)	0.093	-0.16	(0.12)	0.16	+0.22	(0.16)	0.19	-0.22	(0.13)	0.092
Cluster 4: Orange-Blue	-0.08	(0.06)	0.19	-0.08	(0.06)	0.17	-0.10	(0.06)	0.11	-0.05	(0.10)	0.64	-0.10	(0.08)	0.21
All-cause dementia incidence															
Cluster 1: Orange-Red	-0.05	(0.06)	0.37	-0.03	(0.06)	0.64	-0.04	(0.06)	0.53	+0.02	(0.10)	0.86	-0.11	(0.08)	0.15
Cluster 2: Red-Green	+0.12	(0.07)	0.11	+0.12	(0.07)	0.083	+0.15	(0.09)	0.092	-0.15	(0.15)	0.33	+0.21	(0.09)	0.018*
Cluster 3: Yellow-Orange	-0.05	(0.06)	0.44	-0.07	(0.06)	0.22	-0.04	(0.09)	0.64	+0.14	(0.13)	0.25	-0.08	(0.10)	0.43
Cluster 4: Orange-Blue	-0.06	(0.04)	0.18	-0.05	(0.04)	0.21	-0.07	(0.04)	0.64	-0.02	(0.07)	0.79	-0.08	(0.05)	0.16

See Table 1 for periodontal pathogen abbreviations. AD, Alzheimer's disease. ^aModels were adjusted for age, sex, race/ethnicity, poverty income ratio, education (years), urban-rural area of residence, household size, marital status, nutritional factors (HEI, MAR), nutritional biomarkers (25(OH)D, folate, vitamin C, vitamin A, total carotenoids, vitamin E, ferritin, selenium and normalized calcium), lifestyle (smoking, drug use, alcohol, physical activity), health-related factors (self-rated health, co-morbidity index, allostatic load, weight status), dentate status and social support variables, as well as Phase of NHANES III. Covariates (other than exposures) were imputed and analysis is across 5 imputations with 10 iterations. ^b19 Periodontal pathogen exposures (both phases) were Log_e transformed and then standardized into z-scores. Factor analysis was conducted from which 5 factors were extracted each explaining >4% of total variance. After varimax rotation, factor 1 loaded highest ($\lambda > 0.40$) on *T. forsythia* (0.68), *A. actinomycetemcomitans* (0.65), *F. nucleatum* (0.53), *S. oralis* (0.81), *M. micros* (0.51), *C. rectus* (0.54), *E. corrodens* (0.68), *S. intermedius* (0.63), *V. parvula* (0.64), *S. noxia* (0.62), *T. denticola* (0.63) and *S. mutans* (0.73); factor 2 on *P. gingivalis* (0.42), *P. intermedia* (0.85), *P. nigrescens* (0.84), *F. nucleatum* (0.44), *C. rectus* (0.40), *S. intermedius* (0.42), *C. ochracea* (0.44), *P. melaninogenica* (0.70); factor 3 on *E. nodatum* (0.75), *A. naselundii* (0.76); factor 4 on *P. gingivalis* (0.44) and *C. rectus* (0.42); and factor 5 on *C. ochracea* (0.45) and *S. noxia* (0.41). Factors were labelled based on up to 3 highest loadings, using a shortcut name for each. See Supplementary Figure 1 and methods section for definition of each cluster. ^cUnweighted N = 6,277–6,278, weighted mean follow-up time: 200 months (AD mortality), 197 months (AD incidence), 192 months (all-cause dementia); ^d Unweighted N = 4,681–4,682, weighted mean follow-up time: 177 months (AD mortality), 171 months (AD incidence), 165 months (all-cause dementia); ^e Unweighted N = 3,077, weighted mean follow-up time: 148 months (AD mortality), 141 months (AD incidence), 133 months (all-cause dementia); ^f Unweighted N = 2,924, weighted mean follow-up time: 196 months (AD mortality), 193 months (AD incidence), 189 months (all-cause dementia) ^g Unweighted N = 3,353–3,354, weighted mean follow-up time: 205 months (AD mortality), 200 months (AD incidence), 194 months (all-cause dementia). * $p < 0.033$, marginally significant after correction for multiple testing; ** $p < 0.016$, significant after correction for multiple testing.

428 *Clinical Pd markers and their association with*
 429 *periodontal pathogens and AD/all-cause*
 430 *dementia outcomes*

431 Moreover, *P. gingivalis* IgG, the Orange-Red, Red-
 432 Green and Yellow-Orange clusters, Factors 2 and
 433 4 were independently associated with clinical Pd
 434 markers (AL/PPD) (Supplementary Table 3). Nev-
 435 ertheless, only a marginal association between PPD
 436 and incident AD risk was detected among men and
 437 older individuals upon multiple testing adjustment
 438 (Supplementary Table 4).

439 **DISCUSSION**

440 To our knowledge, this is the first large retro-
 441 spective cohort study to examine the association
 442 between periodontal pathogens (and measures of Pd:
 443 AL/PPD) with AD incidence and mortality and inci-
 444 dent all-cause dementia. Our findings indicated that
 445 IgG against *P. gingivalis*, *P. melaninogenica*, and *C.*
 446 *rectus*, two empirical periodontal pathogen factors,
 447 and two empirical periodontal pathogen clusters as
 448 well as PPD were consistently linked with at least
 449 one of the 3 outcomes among older adults. More-
 450 over, findings with all-cause dementia and not AD
 451 pertained mostly to the outcome of vascular demen-
 452 tia, given that it is the second most common cause of
 453 dementia.

454 Although there are no other studies examin-
 455 ing the association between periodontal pathogens
 456 and incidence of dementia per se, several studies
 457 have examined the relations between periodontal
 458 pathogens and cognitive impairments that could yield
 459 dementia outcomes. Our findings of positive associ-
 460 ations between Pd and periodontal pathogens with
 461 various dementia outcomes mirror previous find-
 462 ings with cognitive outcomes. Specifically, using
 463 NHANES III, a study found that the highest Pg
 464 IgG (119 ELISA Units [EU]) were more likely to
 465 exhibit poor delayed verbal recall (OR 2.89, 95%
 466 CI 1.14 to 7.29) and impaired subtraction (OR
 467 1.95, 95% CI 1.22 to 3.11) [13]. Two other nested
 468 case-control studies of periodontal pathogens found
 469 that participants with elevated *A. naeslundii* IgG
 470 (0.640 ng/ml) level exhibited higher risk of AD [14],
 471 as did titers for *F. nucleatum* and *P. intermedia* [15].
 472 Similarly, the risk of developing dementia was higher
 473 among Pd patients compared to controls (HR = 1.16,
 474 95% CI = 1.01–1.32, $p = 0.03$) [33]. Nevertheless,
 475 recent reviews and meta-analyses examining pooled
 476 evidence on Pd and dementia came to different con-

477 clusions [12]. This discrepancy highlights the need to
 478 examine associations between periodontal pathogens
 479 with AD and other types of dementia within different
 480 sub-groups (preferably at different baseline ages), as
 481 was done in our present study. Furthermore, our study
 482 indicated that PPD, a measure of current periodon-
 483 titis was associated with incident AD among older
 484 adults, though that was not the case of AL, a mea-
 485 sure of cumulative exposure. This finding needs to
 486 be replicated in other comparable cohorts.

487 Suggested mechanisms linking Pd or periodontal
 488 pathogens with cognitive impairment and demen-
 489 tia are still speculative. First, bacterial pathogens
 490 can spread from periodontal regions to blood stream
 491 into other bodily organs. Second, toxins produced
 492 by pathogens can damage the vascular system via
 493 oxidative stress leading to atherosclerosis which may
 494 trigger dementia or stroke [12]. Third, inflammatory
 495 mediators of Pd including cytokines, chemokines,
 496 and prostaglandins can contribute to AD by trig-
 497 gering brain inflammation [19]. *P. gingivalis* and
 498 *P. melaninogenica* are related rod-shaped, black-
 499 pigmented, strictly anaerobic gram-negative bacteria
 500 [18]. Perhaps the most characteristic feature of *P.*
 501 *gingivalis* induced periodontitis is the production of
 502 gingipains, enzymes that can cleave proteins specifi-
 503 cally after arginine or lysine amino acids [34], and
 504 which are secreted through a complex known as
 505 Type IX Secretion System (T9SS) protein secre-
 506 tion system [34]. Gingipains target host peptides
 507 with antimicrobial or anti-inflammatory activities,
 508 and by inactivating them induce edema and bleed-
 509 ing, in addition to allowing bacterial cells to infiltrate
 510 neutrophils [35]. Together with other virulence fac-
 511 tors, this allows *P. gingivalis* to induce inflammation
 512 while evading host immune response [36], and to
 513 make use of inflammatory fluids as a source of
 514 essential nutrients (e.g., iron) [37] required for bac-
 515 terial growth [38]. *P. gingivalis* produces proteolytic
 516 enzymes that target immunoglobulins and cell surface
 517 adhesion proteins, which could facilitate invasion and
 518 weaken host immune response. Mouse models show that
 519 the lipopolysaccharides (LPS) and the gingipains pro-
 520 duced by Pg respectively increase accumulation of
 521 amyloid- β (A β) [20, 39] and enhance migration and
 522 inflammation of microglia [40], which are two hall-
 523 mark pathologies of AD. Recent mouse studies have
 524 demonstrated that repeated exposure to *P. gingivalis*,
 525 resulting in gingipain accumulation in and around
 526 brain cells, was responsible for neurodegeneration
 527 and strongly correlated with hippocampal A β accu-
 528 mulation [20, 41] onset [42]. Importantly, Poole et al.

529 confirmed in an *in vitro* study of AD brain tissue and
530 controls that that LPS from periodontal bacteria can
531 access the AD brain during life given that labeling
532 in the matched controls was absent. This demon-
533 stration of a known chronic oral-pathogen-related
534 virulence factor reaching the human brains suggests
535 an inflammatory role in the existing AD pathology
536 [43]. Moreover, Dominy et. al have found that small-
537 molecule inhibitors of gingipains may be an effective
538 treatment against *P. gingivalis*-induced brain inflam-
539 mation and bacterial colonization and thus may slow
540 neurodegeneration [20]. The present study adds to
541 the epidemiological evidence suggesting that *P. gin-*
542 *givalis* eradication among others may be an effective
543 means to delay onset of AD, pending randomized
544 clinical trials.

545 Just like *P. gingivalis*, *P. melaninogenica* expresses
546 a complete T9SS secretion system. Although *P.*
547 *melaninogenica* does not use gingipains, T9SS is
548 important in biofilm formation and is involved in
549 Pd, possibly by secreting proteases [44]. Our results
550 indicate that Pg and Pm may independently or inter-
551 actively induce cognitive impairment leading to AD
552 as an underlying cause of death among older adults.

553 Moreover, another study showed that *P. gingivalis*
554 LPS alone was sufficient to antagonize IL-6 and IL-
555 8, but not IL-1 β stimulation by another pathogen,
556 namely *C. rectus*, suggesting that mixed infections,
557 particularly interactions between *P. gingivalis* and *C.*
558 *rectus* may impair host immune responses through
559 cytokine level reduction of direct relevance to both
560 periodontitis and AD [45].

561 Our study has several notable strengths including
562 the use of a large, nationally representative sample,
563 inclusion of middle-aged adults (≥ 45 years), assess-
564 ment of AD incidence and mortality and incident
565 all-cause dementia over a long follow-up period of
566 up to 26 years, measurement of serum antibody lev-
567 els for periodontal pathogens combined with dental
568 examination and adjustment for key potential con-
569 founders.

570 Limitations include observational study design,
571 even though temporality of associations were ascer-
572 tained. Underdiagnosis of AD and other dementias
573 is a possibility despite the fact that over 90% of
574 the US population is eligible for and uses Medi-
575 care after the age of 65 years and that the linkage
576 was comprehensive, including all aspects of health
577 care utilization (e.g., inpatient and outpatient) with
578 continuous follow-up between 1991 and 2014. Nev-
579 ertheless, a few cases missed by Medicare were
580 added using NDI to assess incident AD and all-

581 cause dementia. Moreover, the data lacked some
582 key biochemical biomarkers of AD (such as blood
583 or CSF markers of A β and tau) and neuroimaging
584 of patients. Additionally, clinical periodontal mea-
585 sures were only estimated based on partial-mouth
586 examination. This could underestimate Pd severity,
587 thus attenuating observed associations. An in-depth
588 study examining other alternative measures of clin-
589 ically defined categories for periodontitis may be
590 warranted. Furthermore, serum IgG humoral immune
591 response exposures, though normalized through
592 Log_e transformation, exhibited moderate collinear-
593 ity. Finally, residual confounding bias particularly by
594 genetic risk factors (e.g., ApoE4 status) cannot be
595 discounted.

596 This study provides evidence for an association
597 between periodontal pathogens and AD, which was
598 stronger for older adults and calls for a line of inquiry,
599 including randomized controlled trials, on the effec-
600 tiveness of periodontal treatment against onset and
601 progression of neurodegenerative disorders such as
602 AD.

603 ACKNOWLEDGMENTS

604 The authors would like to thank Mr. Negasi Beyene
605 (CDC/NCHS/RDC) and Mr. Ray Kuntz (AHRQ) for
606 their help with access to the restricted data at the
607 RDC and facilitating data analysis at the AHRQ
608 headquarters, Rockville, MD. We would also like
609 to thank Ms. Megan Williams and Ms. Nicolle
610 Mode (NIA/NIH/IRP) for internally reviewing our
611 manuscript.

612 This study was entirely supported by the National
613 Institute on Aging, Intramural Research Program
614 (NIA/NIH/IRP).

615 The views expressed in this article are those of
616 the author(s) and do not reflect the official policy of
617 the Department of the Army/Navy/Air Force, Depart-
618 ment of Defense, or the U.S. Government.

619 Authors' disclosures available online ([https://](https://www.j-alz.com/manuscript-disclosures/20-0064r1)
620 www.j-alz.com/manuscript-disclosures/20-0064r1).

621 SUPPLEMENTARY MATERIAL

622 The supplementary material is available in the
623 electronic version of this article: [http://dx.doi.org/10.](http://dx.doi.org/10.3233/JAD-200064)
624 [3233/JAD-200064](http://dx.doi.org/10.3233/JAD-200064).

REFERENCES

- 624
- 625 [1] Sosa-Ortiz AL, Acosta-Castillo I, Prince MJ (2012) Epidemiology of dementias and Alzheimer's disease. *Arch Med Res* **43**, 600-608. 688
- 626
- 627
- 628 [2] Ferri CP, Prince M, Brayne C, Brodaty H, Fratiglioni L, Ganguli M, Hall K, Hasegawa K, Hendrie H, Huang Y, Jorm A, Mathers C, Menezes PR, Rimmer E, Sczufca M, Alzheimer's Disease International (2005) Global prevalence of dementia: A Delphi consensus study. *Lancet* **366**, 2112-2117. 689
- 629
- 630
- 631
- 632
- 633
- 634 [3] Prince M, Bryce R, Albanese E, Wimo A, Ribeiro W, Ferri CP (2013) The global prevalence of dementia: A systematic review and metaanalysis. *Alzheimers Dement* **9**, 63-75 e62. 690
- 635
- 636
- 637 [4] Lindeboom J, Weinstein H (2004) Neuropsychology of cognitive ageing, minimal cognitive impairment, Alzheimer's disease, and vascular cognitive impairment. *Eur J Pharmacol* **490**, 83-86. 691
- 638
- 639
- 640
- 641 [5] Hardy J, Selkoe DJ (2002) The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science* **297**, 353-356. 692
- 642
- 643
- 644 [6] Turner RS (2003) Biomarkers of Alzheimer's disease and mild cognitive impairment: Are we there yet? *Exp Neurol* **183**, 7-10. 693
- 645
- 646
- 647 [7] Helmer C, Pasquier F, Dartigues JF (2006) [Epidemiology of Alzheimer disease and related disorders]. *Med Sci (Paris)* **22**, 288-296. 694
- 648
- 649
- 650 [8] Honjo K, van Reekum R, Verhoeff NP (2009) Alzheimer's disease and infection: Do infectious agents contribute to progression of Alzheimer's disease? *Alzheimers Dement* **5**, 348-360. 695
- 651
- 652
- 653
- 654 [9] Alzheimer's Association (2016) 2016 Alzheimer's disease facts and figures. *Alzheimers Dement* **12**, 459-509. 696
- 655
- 656 [10] Barnes DE, Yaffe K (2011) The projected effect of risk factor reduction on Alzheimer's disease prevalence. *Lancet Neurol* **10**, 819-828. 697
- 657
- 658
- 659 [11] Cestari JA, Fabri GM, Kalil J, Nitrini R, Jacob-Filho W, de Siqueira JT, Siqueira SR (2016) Oral infections and cytokine levels in patients with Alzheimer's disease and mild cognitive impairment compared with controls. *J Alzheimers Dis* **52**, 1479-1485. 698
- 660
- 661
- 662
- 663
- 664 [12] Maldonado A, Laugisch O, Burgin W, Sculean A, Eick S (2018) Clinical-periodontal variables in patients with and without dementia-a systematic review and meta-analysis. *Clin Oral Investig* **22**, 2463-2474. 699
- 665
- 666
- 667 [13] Noble JM, Borrell LN, Papapanou PN, Elkind MS, Scarmeas N, Wright CB (2009) Periodontitis is associated with cognitive impairment among older adults: Analysis of NHANES-III. *J Neurol Neurosurg Psychiatry* **80**, 1206-1211. 700
- 668
- 669
- 670
- 671
- 672
- 673 [14] Noble JM, Scarmeas N, Celenti RS, Elkind MS, Wright CB, Schupf N, Papapanou PN (2014) Serum IgG antibody levels to periodontal microbiota are associated with incident Alzheimer disease. *PLoS One* **9**, e114959. 701
- 674
- 675
- 676
- 677 [15] Sparks Stein P, Steffen MJ, Smith C, Jicha G, Ebersole JL, Abner E, Dawson D, 3rd (2012) Serum antibodies to periodontal pathogens are a risk factor for Alzheimer's disease. *Alzheimers Dement* **8**, 196-203. 702
- 678
- 679
- 680
- 681 [16] Leira Y, Dominguez C, Seoane J, Seoane-Romero J, Pias-Peleiteiro JM, Takkouche B, Blanco J, Aldrey JM (2017) Is periodontal disease associated with Alzheimer's disease? A systematic review with meta-analysis. *Neuroepidemiology* **48**, 21-31. 703
- 682
- 683
- 684
- 685
- 686 [17] Pihlstrom BL, Michalowicz BS, Johnson NW (2005) Periodontal diseases. *Lancet* **366**, 1809-1820. 704
- 687
- 688 [18] Kamer AR, Dasanayake AP, Craig RG, Glodzik-Sobanska L, Bry M, de Leon MJ (2008) Alzheimer's disease and peripheral infections: The possible contribution from periodontal infections, model and hypothesis. *J Alzheimers Dis* **13**, 437-449. 705
- 689
- 690
- 691
- 692
- 693 [19] Kamer AR, Craig RG, Dasanayake AP, Brys M, Glodzik-Sobanska L, de Leon MJ (2008) Inflammation and Alzheimer's disease: Possible role of periodontal diseases. *Alzheimers Dement* **4**, 242-250. 706
- 694
- 695
- 696
- 697 [20] Dominy SS, Lynch C, Ermini F, Benedyk M, Marczyk A, Konradi A, Nguyen M, Haditsch U, Raha D, Griffin C, Holsinger LJ, Arastu-Kapur S, Kaba S, Lee A, Ryder MI, Potempa B, Mydel P, Hellvard A, Adamowicz K, Hasturk H, Walker GD, Reynolds EC, Faull RLM, Curtis MA, Dragunow M, Potempa J (2019) Porphyromonas gingivalis in Alzheimer's disease brains: Evidence for disease causation and treatment with small-molecule inhibitors. *Sci Adv* **5**, eaau3333. 707
- 698
- 699
- 700
- 701 [21] Center for Disease Control and Prevention (CDC) (1996) Centers for Disease Control and Prevention. 708
- 702
- 703 [22] Center for Disease Control and Prevention, NHANES and CMS Linked Data Overview, <https://www.cdc.gov/nchs/tutorials/NHANES-CMS/Orientation/Overview/index.htm>. 709
- 704
- 705
- 706 [23] National Center for Health Statistics, Data Linkage (2015) Underlying and Multiple Cause of Death Codes. URL: https://www.cdc.gov/nchs/nvss/mortality_public_use_data.htm. 710
- 707
- 708
- 709
- 710 [24] Choi YH, McKeown RE, Mayer-Davis EJ, Liese AD, Song KB, Merchant AT (2011) Association between periodontitis and impaired fasting glucose and diabetes. *Diabetes Care* **34**, 381-386. 711
- 711
- 712 [25] (1994) Plan and operation of the Third National Health and Nutrition Examination Survey, 1988-94. Series 1: Programs and collection procedures. *Vital Health Stat* **1**, 1-407. 712
- 713
- 714 [26] Choi YH, McKeown RE, Mayer-Davis EJ, Liese AD, Song KB, Merchant AT (2014) Serum C-reactive protein and immunoglobulin G antibodies to periodontal pathogens may be effect modifiers of periodontitis and hyperglycemia. *J Periodontol* **85**, 1172-1181. 713
- 715
- 716 [27] Papapanou PN, Neiderud AM, Sandros J, Dahlen G (2001) Checkerboard assessments of serum antibodies to oral microbiota as surrogate markers of clinical periodontal status. *J Clin Periodontol* **28**, 103-106. 714
- 717
- 718 [28] Shrestha D, Choi YH, Zhang J, Hazlett LJ, Merchant AT (2015) Relationship between serologic markers of periodontal bacteria and metabolic syndrome and its components. *J Periodontol* **86**, 418-430. 715
- 719
- 720 [29] Desvarieux M, Demmer RT, Jacobs DR Jr., Rundek T, Boden-Albala B, Sacco RL, Papapanou PN (2010) Periodontal bacteria and hypertension: The oral infections and vascular disease epidemiology study (INVEST). *J Hypertens* **28**, 1413-1421. 716
- 721
- 722 [30] STATA (2017) Statistics/Data Analysis: Release 15.0. Stata Corporation, Texas. URL: <http://www.stata.com>. 717
- 723
- 724 [31] Lee KJ, Carlin JB (2010) Multiple imputation for missing data: Fully conditional specification versus multivariate normal imputation. *Am J Epidemiol* **171**, 624-632. 718
- 725
- 726 [32] Hochberg Y, Tamhane AC (1987) *Multiple comparison procedures*. Wiley, New York. 719
- 727
- 728 [33] Lee YT, Lee HC, Hu CJ, Huang LK, Chao SP, Lin CP, Su EC, Lee YC, Chen CC (2017) Periodontitis as a modifiable risk factor for dementia: A nationwide population-based cohort study. *J Am Geriatr Soc* **65**, 301-305. 720
- 729
- 730
- 731
- 732
- 733
- 734
- 735
- 736
- 737
- 738
- 739
- 740
- 741
- 742
- 743
- 744
- 745
- 746
- 747
- 748
- 749
- 750
- 751
- 752

- 753 [34] Potempa J, Sroka A, Imamura T, Travis J (2003) Gingi- 781
754 pains, the major cysteine proteinases and virulence factors of 782
755 Porphyromonas gingivalis: Structure, function and assembly 783
756 of multidomain protein complexes. *Curr Protein Pept* 784
757 *Sci* **4**, 397-407. 785
- 758 [35] Sochalska M, Potempa J (2017) Manipulation of neutrophils 786
759 by Porphyromonas gingivalis in the development of peri- 787
760 odontitis. *Front Cell Infect Microbiol* **7**, 197. 788
- 761 [36] Tada H, Nishioka T, Takase A, Numazaki K, Bando K, 789
762 Matsushita K (2019) Porphyromonas gingivalis induces the 790
763 production of interleukin-31 by human mast cells, result- 791
764 ing in dysfunction of the gingival epithelial barrier. *Cell* 792
765 *Microbiol* **21**, e12972. 793
- 766 [37] Hajishengallis G (2011) Immune evasion strategies of Por- 794
767 phyromonas gingivalis. *J Oral Biosci* **53**, 233-240. 795
- 768 [38] Olczak T, Simpson W, Liu X, Genco CA (2005) Iron 796
769 and heme utilization in Porphyromonas gingivalis. *FEMS* 797
770 *Microbiol Rev* **29**, 119-144. 798
- 771 [39] Ishida N, Ishihara Y, Ishida K, Tada H, Funaki-Kato Y, Hagi- 799
772 wara M, Ferdous T, Abdullah M, Mitani A, Michikawa 800
773 M, Matsushita K (2017) Periodontitis induced by bacte- 801
774 rial infection exacerbates features of Alzheimer's disease 802
775 in transgenic mice. *NPJ Aging Mech Dis* **3**, 15. 803
- 776 [40] Liu Y, Wu Z, Nakanishi Y, Ni J, Hayashi Y, Takayama 804
777 F, Zhou Y, Kadowaki T, Nakanishi H (2017) Infection 805
778 of microglia with Porphyromonas gingivalis promotes 806
779 cell migration and an inflammatory response through 807
780 the gingipain-mediated activation of protease-activated 808
receptor-2 in mice. *Sci Rep* **7**, 11759. 809
- [41] Ilievski V, Zuchowska PK, Green SJ, Toth PT, Ragozzino 781
ME, Le K, Aljewari HW, O'Brien-Simpson NM, Reynolds 782
EC, Watanabe K (2018) Chronic oral application of a 783
periodontal pathogen results in brain inflammation, neu- 784
rodegeneration and amyloid beta production in wild type 785
mice. *PLoS One* **13**, e0204941. 786
- [42] Grenier D, Mayrand D, McBride BC (1989) Further studies 787
on the degradation of immunoglobulins by black-pigmented 788
Bacteroides. *Oral Microbiol Immunol* **4**, 12-18. 789
- [43] Poole S, Singhrao SK, Kesavalu L, Curtis MA, Crean S 790
(2013) Determining the presence of periodontopathic viru- 791
lence factors in short-term postmortem Alzheimer's disease 792
brain tissue. *J Alzheimers Dis* **36**, 665-677. 793
- [44] Kondo Y, Sato K, Nagano K, Nishiguchi M, Hoshino T, 794
Fujiwara T, Nakayama K (2018) Involvement of PorK, a 795
component of the type IX secretion system, in Prevotella 796
melaninogenica pathogenicity. *Microbiol Immunol* **62**, 554- 797
566. 798
- [45] Bostanci N, Allaker RP, Belibasakis GN, Rangarajan M, 799
Curtis MA, Hughes FJ, McKay IJ (2007) Porphyromonas 800
gingivalis antagonises Campylobacter rectus induced 801
cytokine production by human monocytes. *Cytokine* **39**, 802
147-156. 803